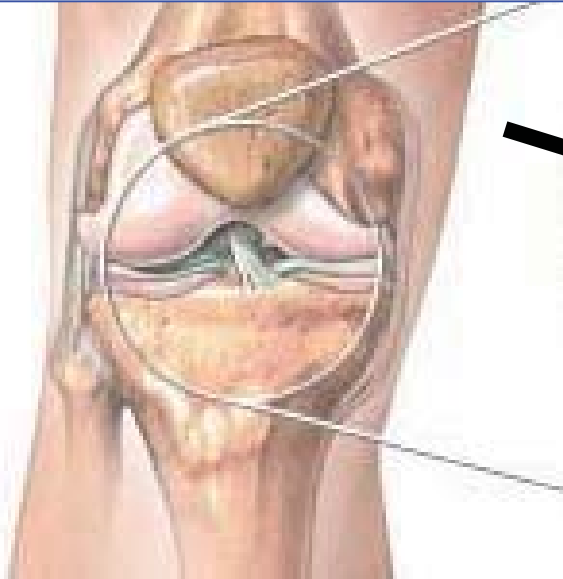
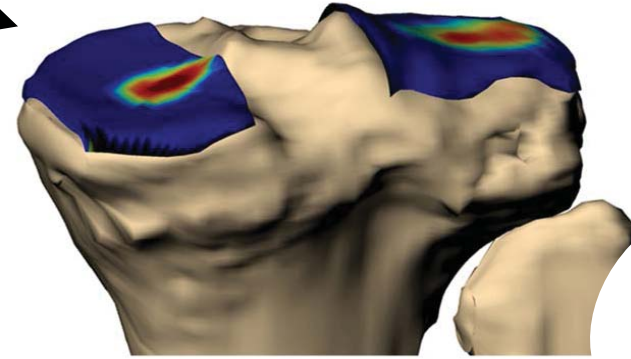


# Mechanics ↔ Biology: Organ, Tissue, Cell, & Molecular Levels

Joint  
(organ)

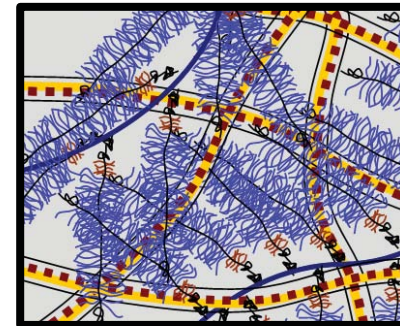
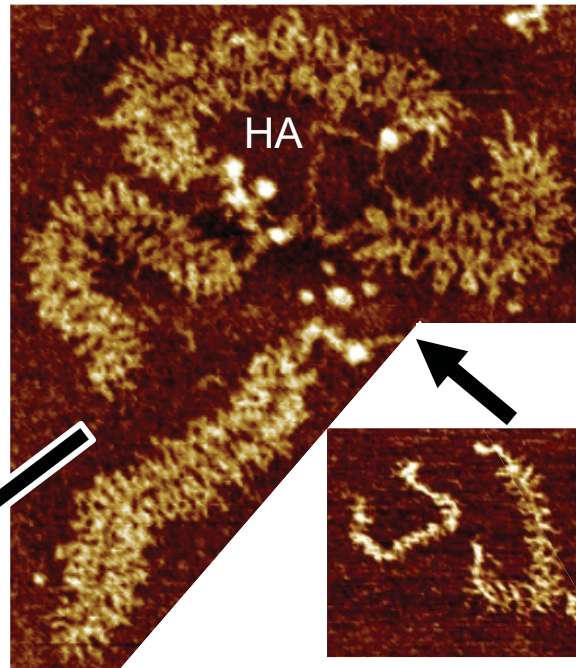
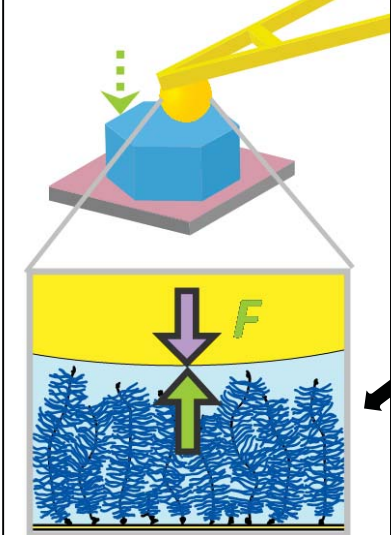


Joint Loading



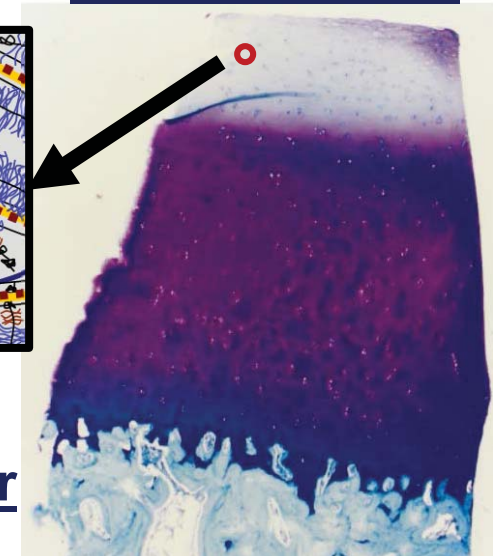
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Molecular  
Mechanics



Extracellular  
Matrix

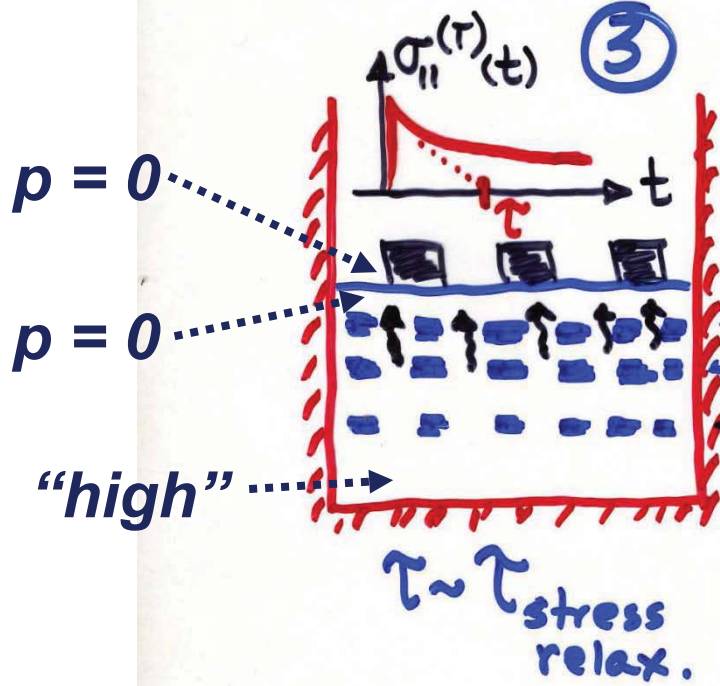
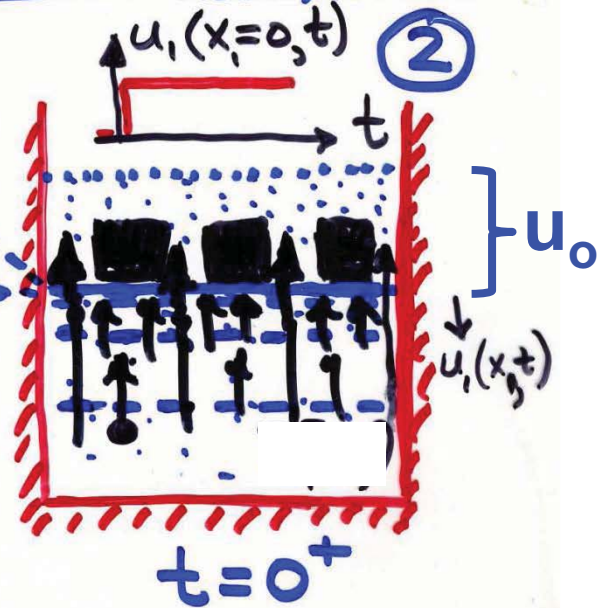
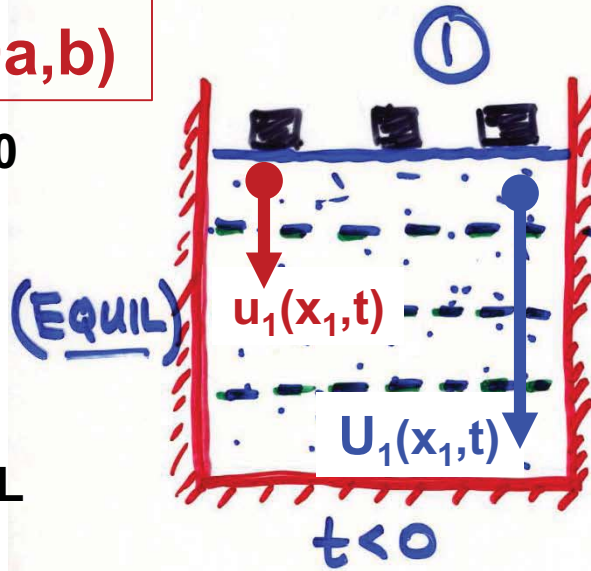
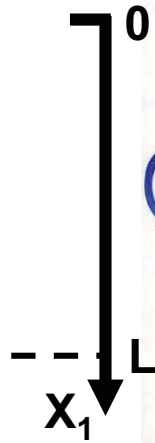
Tissue, Cells



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**PSet 5 #1**  
**(Prob 7.10a,b)**

**POROELASTICITY: STRESS RELAXATION**



$$\tau_{\text{gel}} = \frac{L^2}{\pi^2 D_{\text{gel}}}$$

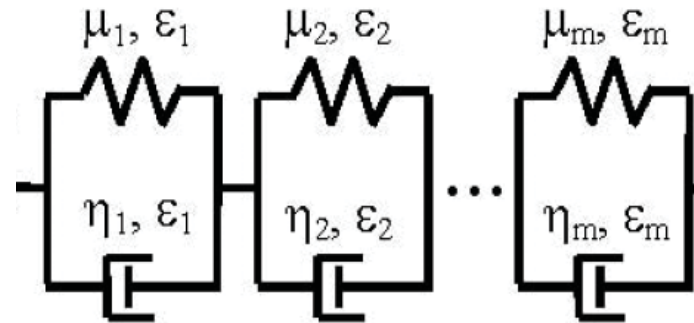
$$D_{\text{gel}} = H \cdot k$$

# Questions:

- What distinguishes solid-phase viscoelasticity..

$$T + \alpha \frac{dT}{dt} = E_1 e + \beta \frac{de}{dt}$$

Time Only.....



....from poroelasticity?

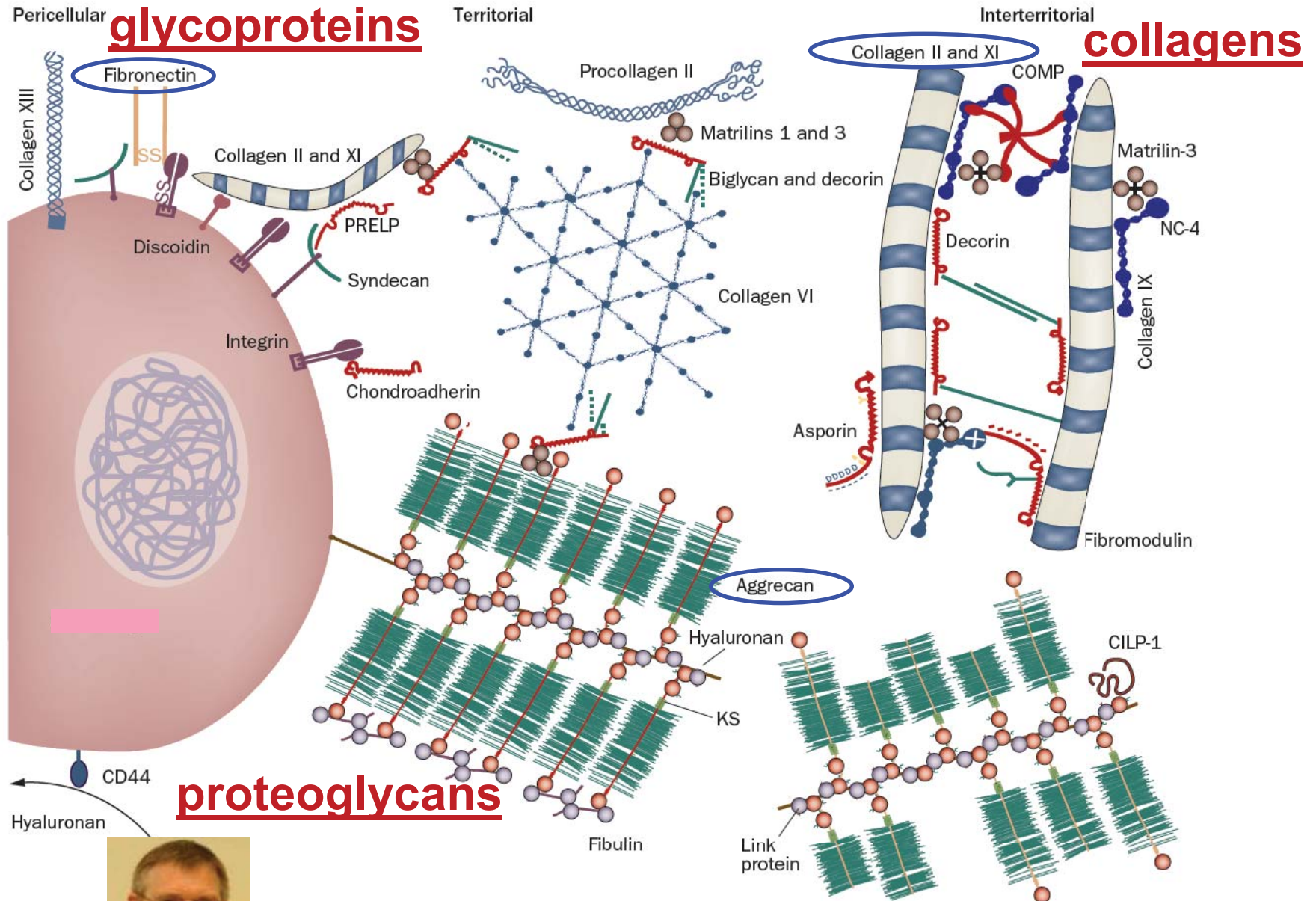


Space  
and Time

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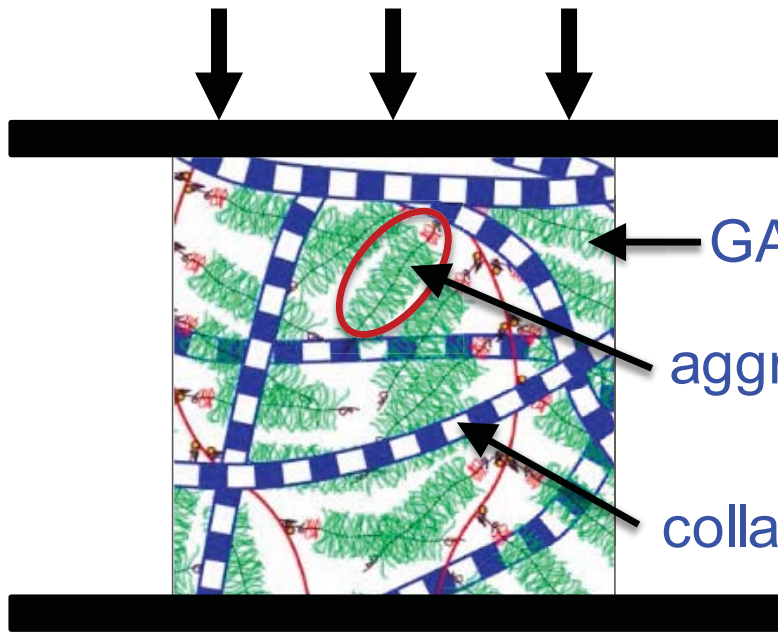
- Does poroelasticity operate at cellular and molecular scales as well as tissue scale??

# Cells Synthesize 100s of Extracellular Matrix Macromolecules



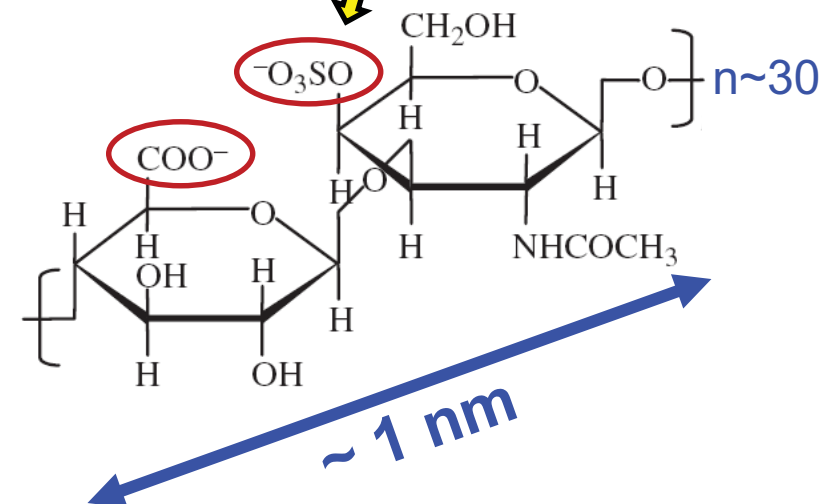
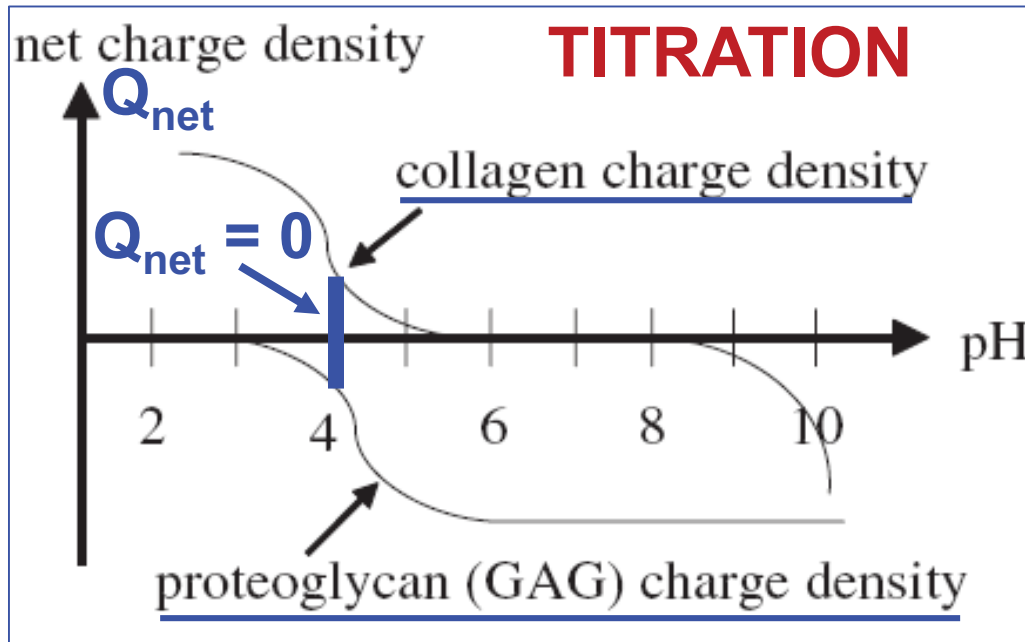
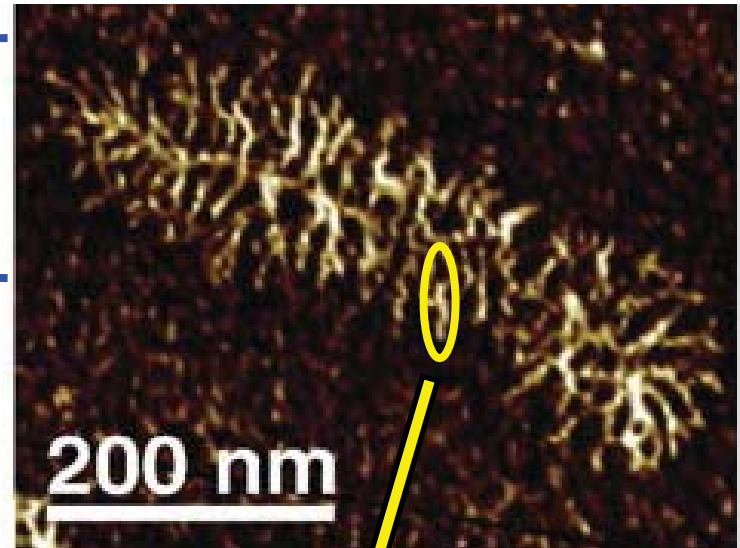
(Dick Heinegård, Nature Revs. Rheumatology 2010)

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 Source: Heinegård, Dick, and Tore Saxne. "The Role of the Cartilage Matrix in Osteoarthritis." *Nature Reviews Rheumatology* 7, no. 1 (2011): 50-56.



$H, k = f(\text{pH, Ionic strength})$   
 [PSet 4, Prob 1]

GAGs  
 aggrecan  
 collagen



# The Amino-acid Composition and Titration Curve of Collagen

BY JOANE H. BOWES AND R. H. KENTEN

*The British Leather Manufacturers' Research Association, London, S.E. 1*

Biochem J

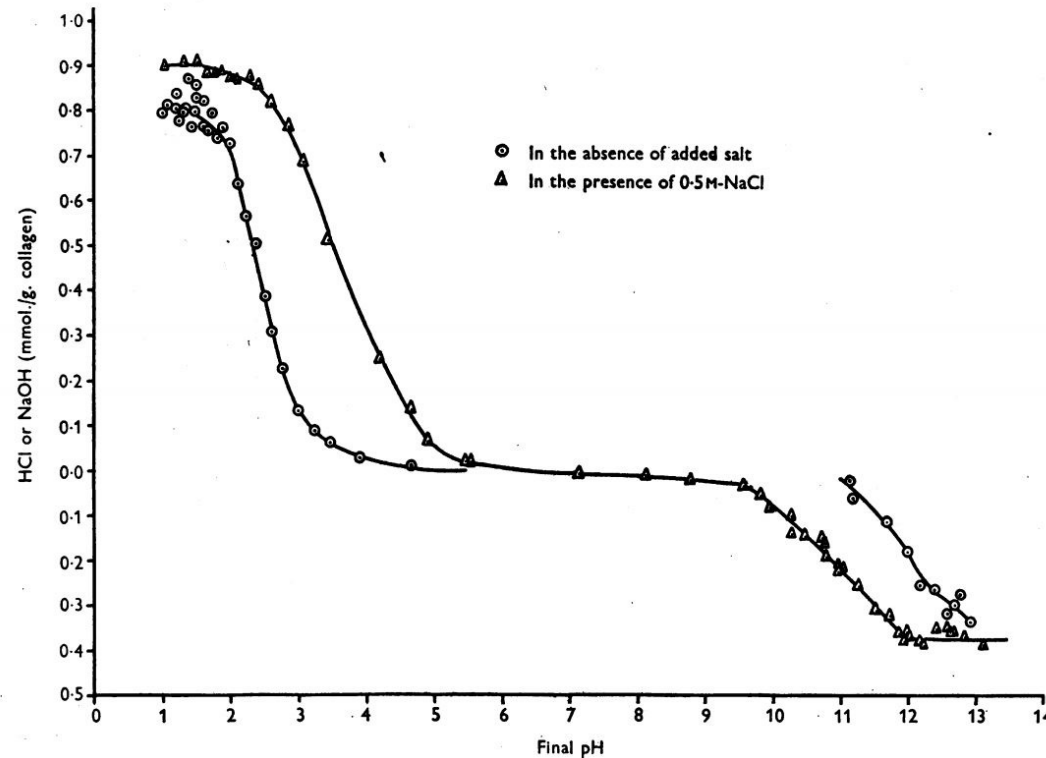
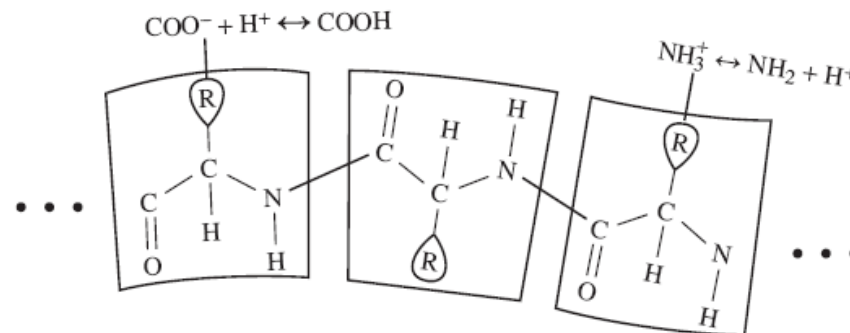


Fig. 2. Titration curves of collagen with and without sodium chloride.

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Source: Bowes, Joane H., and R. H. Kenten. "The Amino-acid Composition and Titration Curve of Collagen." *J. Biol. Chem.*, no. 3 (1948): 358.



- Aggrecan density in tissues subjected to compression (cartilage, disc, tendon) is 10 – 40 X higher than this image (“H”)
- GAGs also resist fluid flow (“k”)

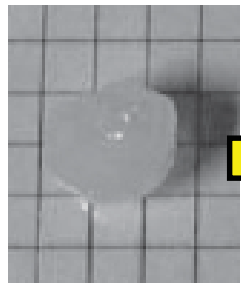


400 nm

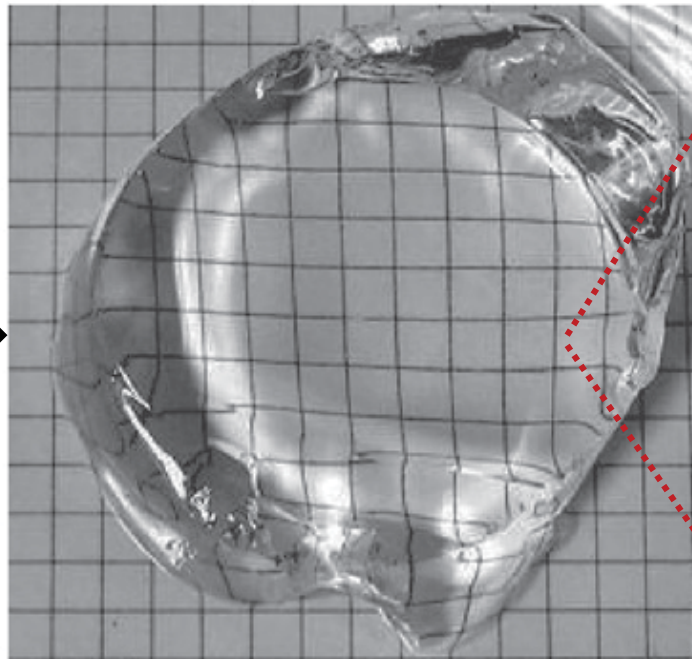
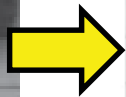
(Fetal bovine aggrecan, Laurel Ng)

# Swelling (“H”) & Fluid Flow (“k”) in "Bio-Porous Media": Molecular Networks & Gels

**Polyelectrolyte Gels Swell:**  
Electrostatic Forces (“H”)

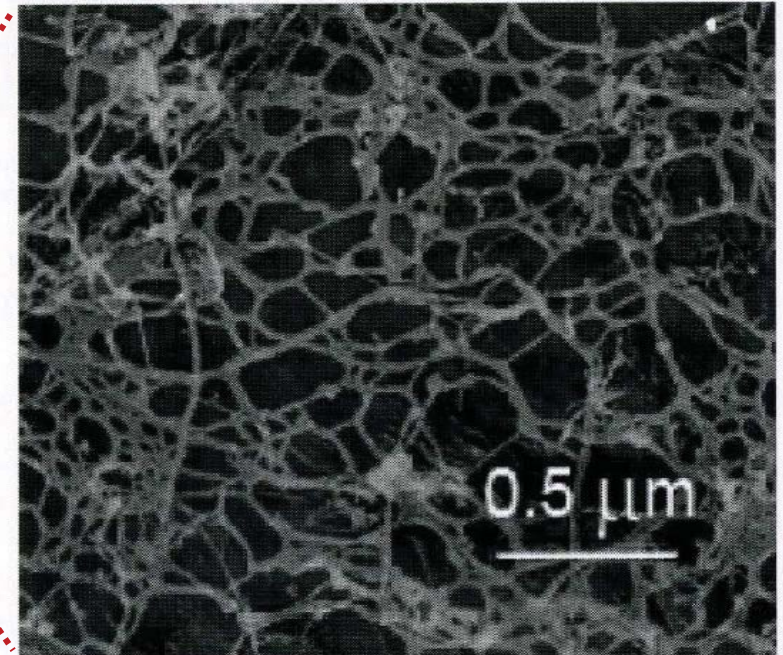


**Neutral**



**Charged**

**Network resists  
fluid flow (“k”)**



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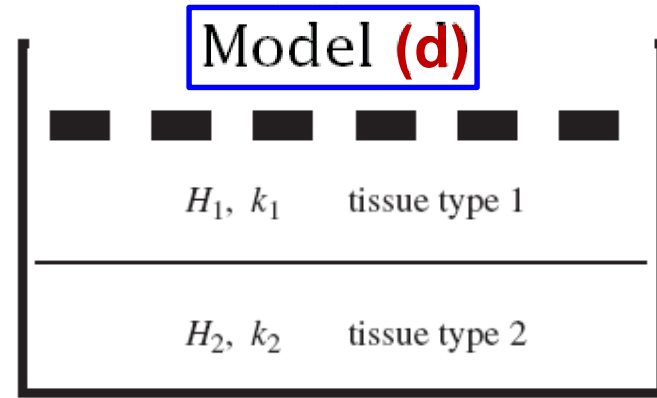
Courtesy of Macmillan Publishers Limited. Used with permission.  
Source: Ono, Toshikazu, et al. "Lipophilic Polyelectrolyte Gels as Super-absorbent Polymers for Nonpolar Organic Solvents." *Nature Materials* 6, no. 6 (2007): 429-33.



Problem 7.11 and Figure 7.34 removed due to copyright restrictions. See the problem in the textbook.  
Source: Grodzinsky, Alan. *Field, Forces and Flows in Biological Systems*. Garland Science, 2011.

Problem 7.11 and Figure 7.34 removed due to copyright restrictions. See the [problem](#) in the textbook.  
Source: Grodzinsky, Alan. *Field, Forces and Flows in Biological Systems*. Garland Science, 2011.

# Two-Layer Poroelastic Model



Problem 7.11 and Figure 7.34 removed due to copyright restrictions. See the problem in the textbook.  
Source: Grodzinsky, Alan. *Field, Forces and Flows in Biological Systems*. Garland Science, 2011.

**PSet 5**  
**Prob 3**

Motivated by:  
Malaviya  
J Orthop Res, 2000

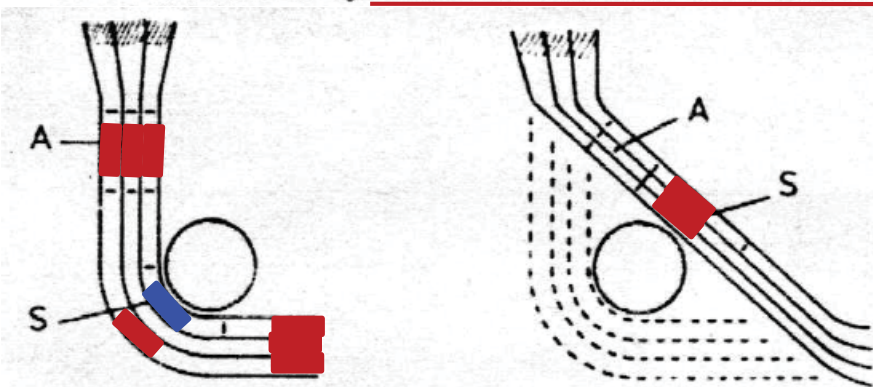
# THE INFLUENCE OF MECHANICAL FORCES ON THE GLYCOSAMINOGLYCAN CONTENT OF THE RABBIT FLEXOR DIGITORUM PROFUNDUS TENDON

GERALD C. GILLARD, HELEN C. REILLY, PAUL G. BELL-BOOTH and MICHAEL H. FLINT

Department of Surgery, School of Medicine, University of Auckland, Private Bag, Auckland 1, New Zealand

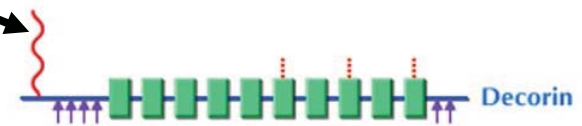


- The physical forces acting on the flexor digitorum profundus tendon of the rabbit were altered by anterior translocation of the tendon. The glycosaminoglycan (GAG) content was determined in regions of the tendon previously under tension or previously subjected to pressure.
- The original pressure bearing region showed a rapid loss of total GAG. This was mainly due to a loss of chondroitin sulfate component, and eventually the region showed a GAG composition similar to that of normal tension transmitting tendon. Replacement of the translocated tendon to its normal position resulted in a slow replacement of the GAG, particularly chondroitin sulfate
- Later when tension was restored to the translocated tendon, the content of these two GAG decreased to normal values while the high overall GAG concentration was maintained by increased amounts of dermatan sulfate.



Chondroitin sulfate ≡ Aggrecan

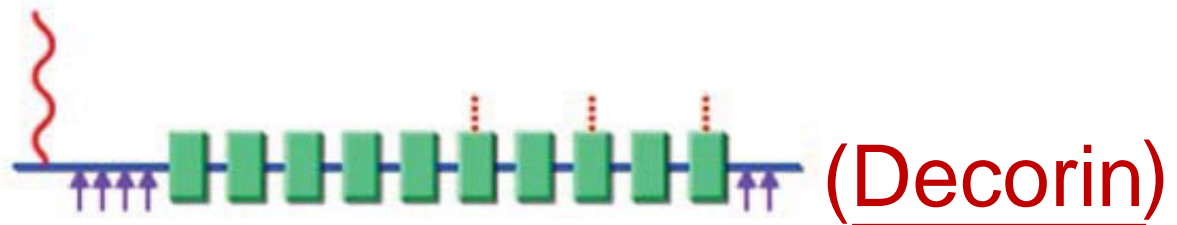
Dermatan sulfate ≡ decorin ("SLRPs")



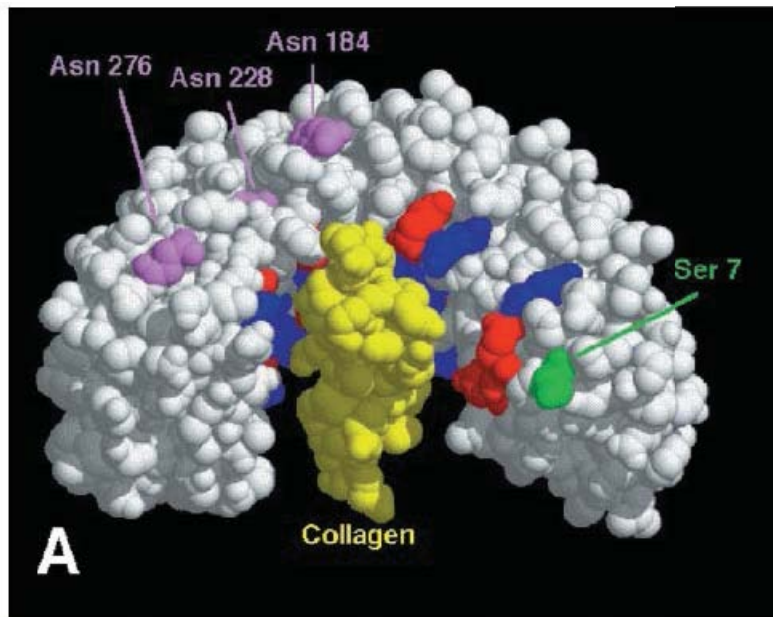
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 Source: Gillard, Gerald C., et al. "The Influence of Mechanical Forces on the Glycosaminoglycan Content of the Rabbit Flexor Digitorum Profundus Tendon." *Connective Tissue Research* 7, no. 1 (1979): 37-46.

# PROTEOGLYCAN SUPERFAMILY

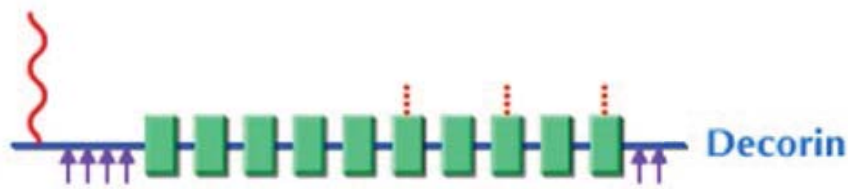
- ECM molecules with (1) Core protein, and (2) Glycosaminoglycan (GAG) chains
- **“Sub-families” of extracellular PGs:**
  - Large Aggregating (Aggrecan)
  - Small Leucine-Rich PG (SLRPs)



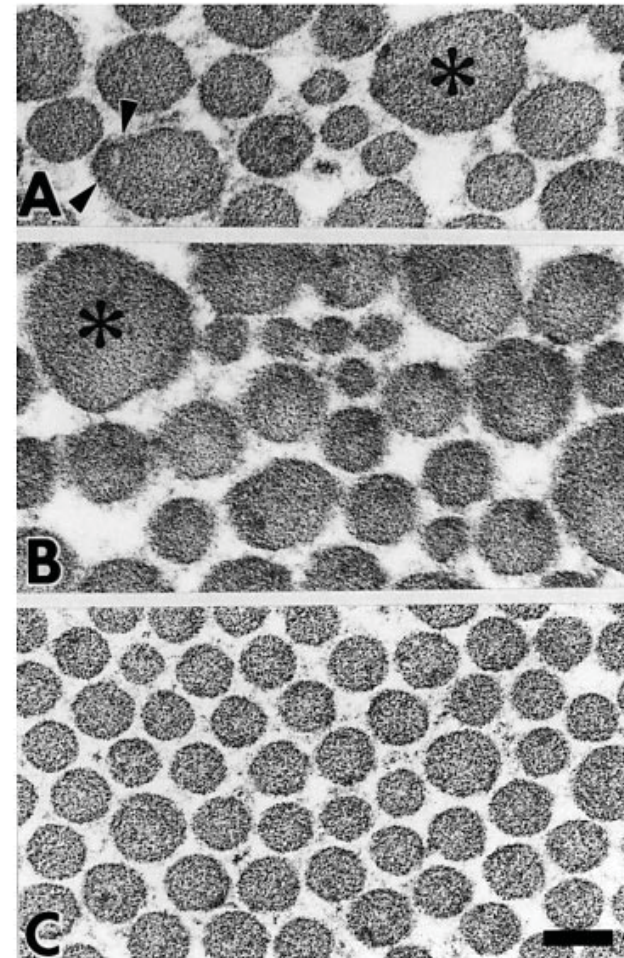
# Decorin & Tendon Collagen Fibrillogenesis



Courtesy of The Journal of Biological Chemistry. Used with permission.  
Source: Iozzo, Renato V. "The Biology of the Small Leucine-rich Proteoglycans Functional Network of Interactive Proteins." *J Biol Chem* 274, no. 27 (1999): 18843-6.



Example of  
Small-Leucine-Rich  
Proteoglycans



SKIN  
decorin  
KO

decorin  
KO

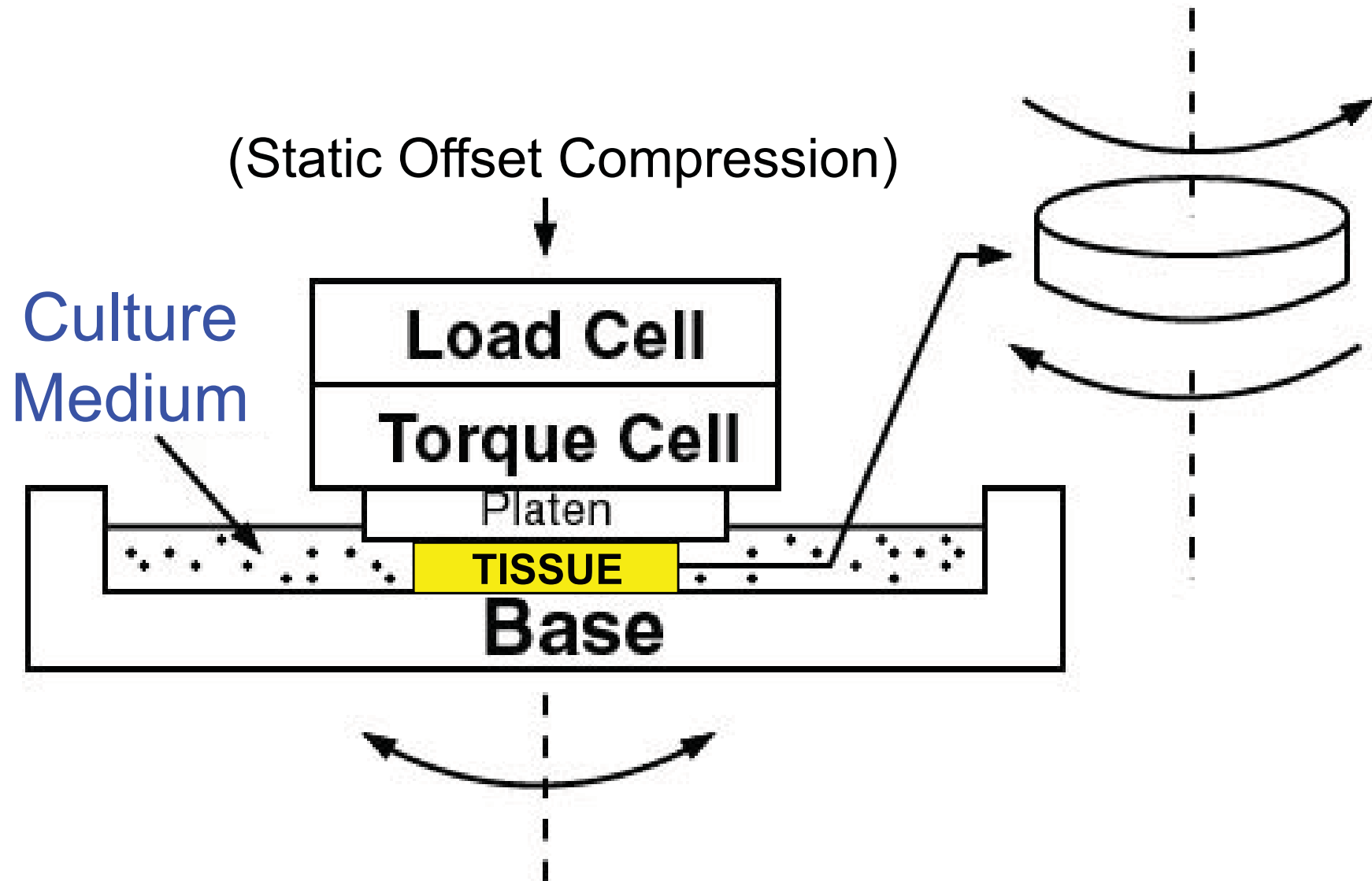
Normal  
(WT)

Figure 4 Ultrastructural appearance of dermal collagen from the skin of decorin null (A and B) and wild-type (C) mice. Notice the larger and irregular cross-sectional profiles in the decorin null collagen fibers (asterisks) with evidence of lateral fusion (A, arrowheads). Bar: 90 nm.

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Source: Iozzo, Renato V. "Matrix Proteoglycans: From Molecular Design to Cellular Function." *J Biol Chem* 273, no. 1 (1998): 609-52.

(Iozzo +, Normal and decorin null mice, J Biol Chem 1999)

# Dynamic Torsional Shear

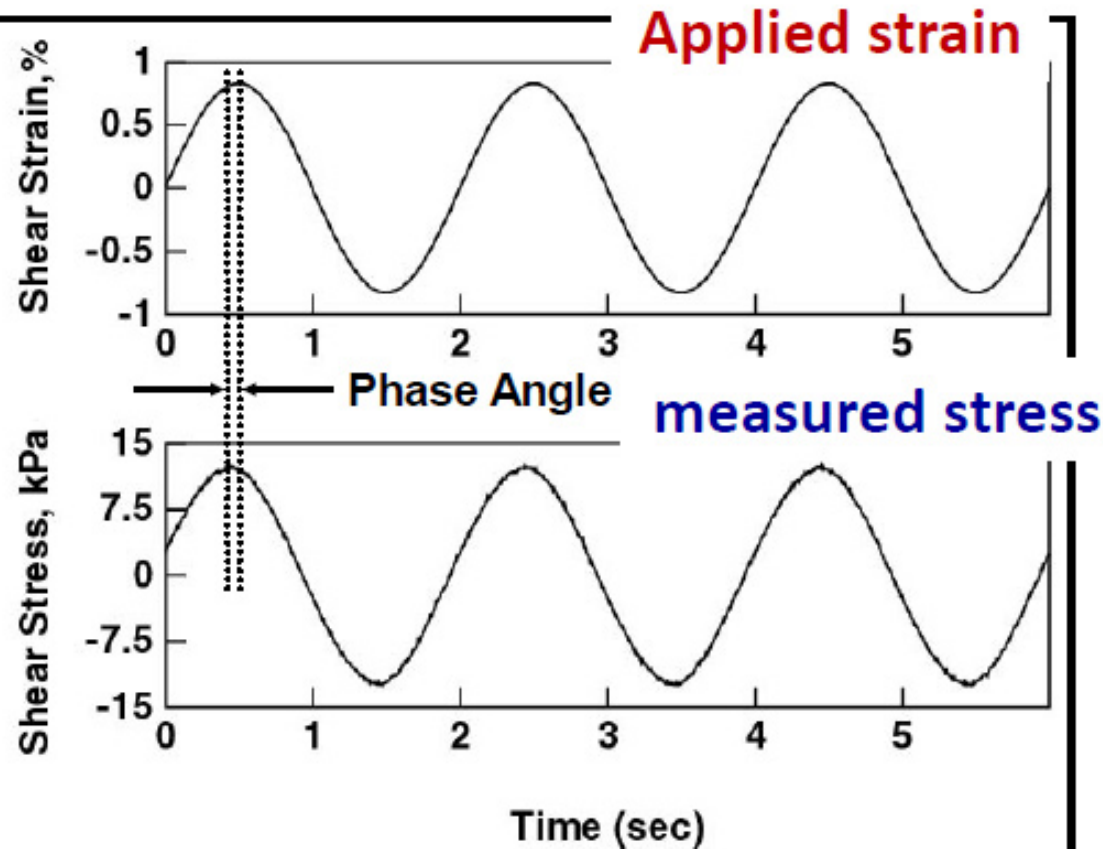
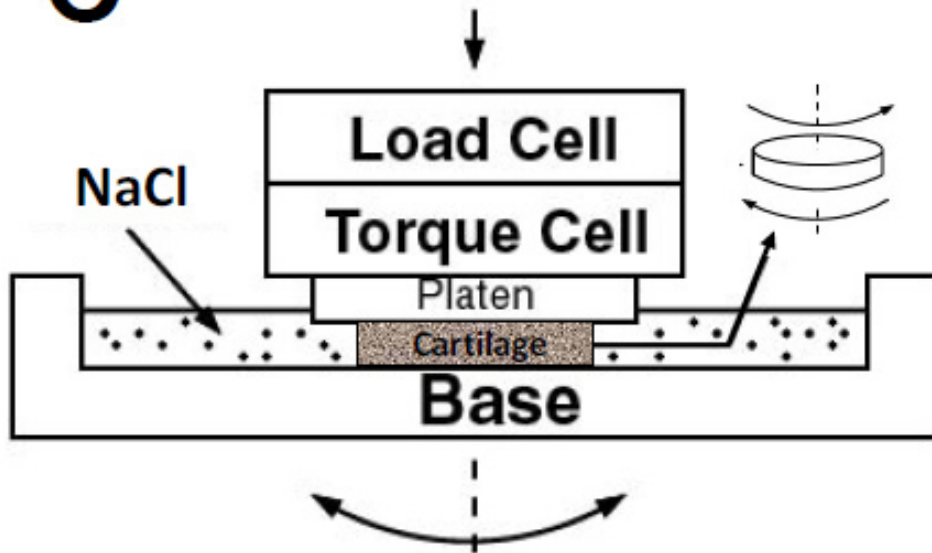


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Source: Jin, Moonsoo, and Alan J. Grodzinsky. "Effect of Electrostatic Interactions between Glycosaminoglycans on the Shear Stiffness of Cartilage: A Molecular Model and Experiments." *Macromolecules* 34, no. 23 (2001): 8330-39.

# “Dynamic Torsional Shear”

Apply sinusoidal shear strain (0.8% amplitude at 0.5 Hz)  
and measure sinusoidal stress amplitude & phase

**C**



Jin+, Macromolecules, 2001

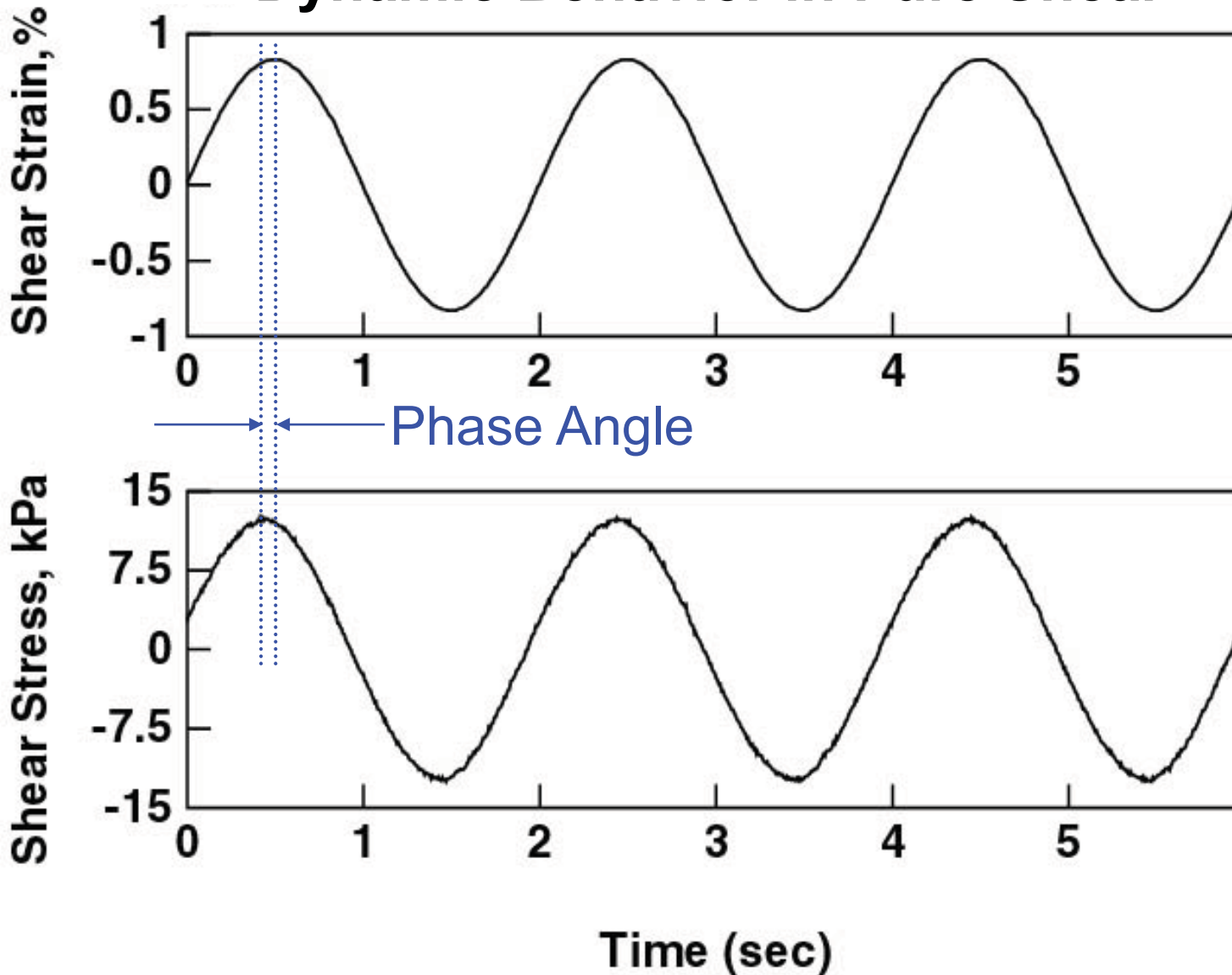
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Source: Jin, Moonsoo, and Alan J. Grodzinsky. "Effect of Electrostatic Interactions between Glycosaminoglycans on the Shear Stiffness of Cartilage: A Molecular Model and Experiments." *Macromolecules* 34, no. 23 (2001): 8330-39.

Is Phase Delay due to  
Viscoelastic **-OR-** Poroelastic behavior?



Non-zero phase angle → Viscoelastic? Poroelastic? both??

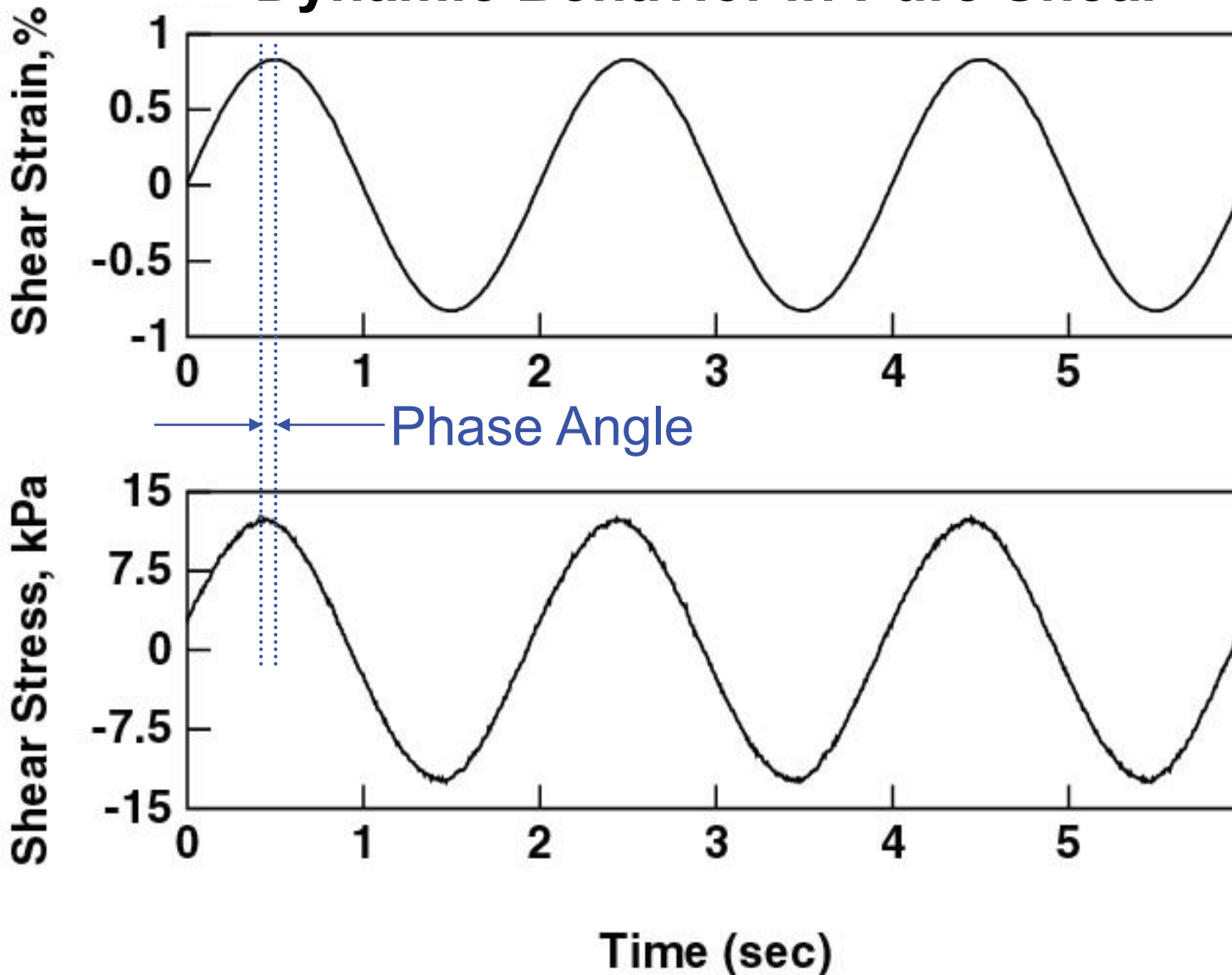
## Dynamic Behavior in Pure Shear



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Source: Jin, Moonsoo, and Alan J. Grodzinsky. "Effect of Electrostatic Interactions between Glycosaminoglycans on the Shear Stiffness of Cartilage: A Molecular Model and Experiments." *Macromolecules* 34, no. 23 (2001): 8330-39.

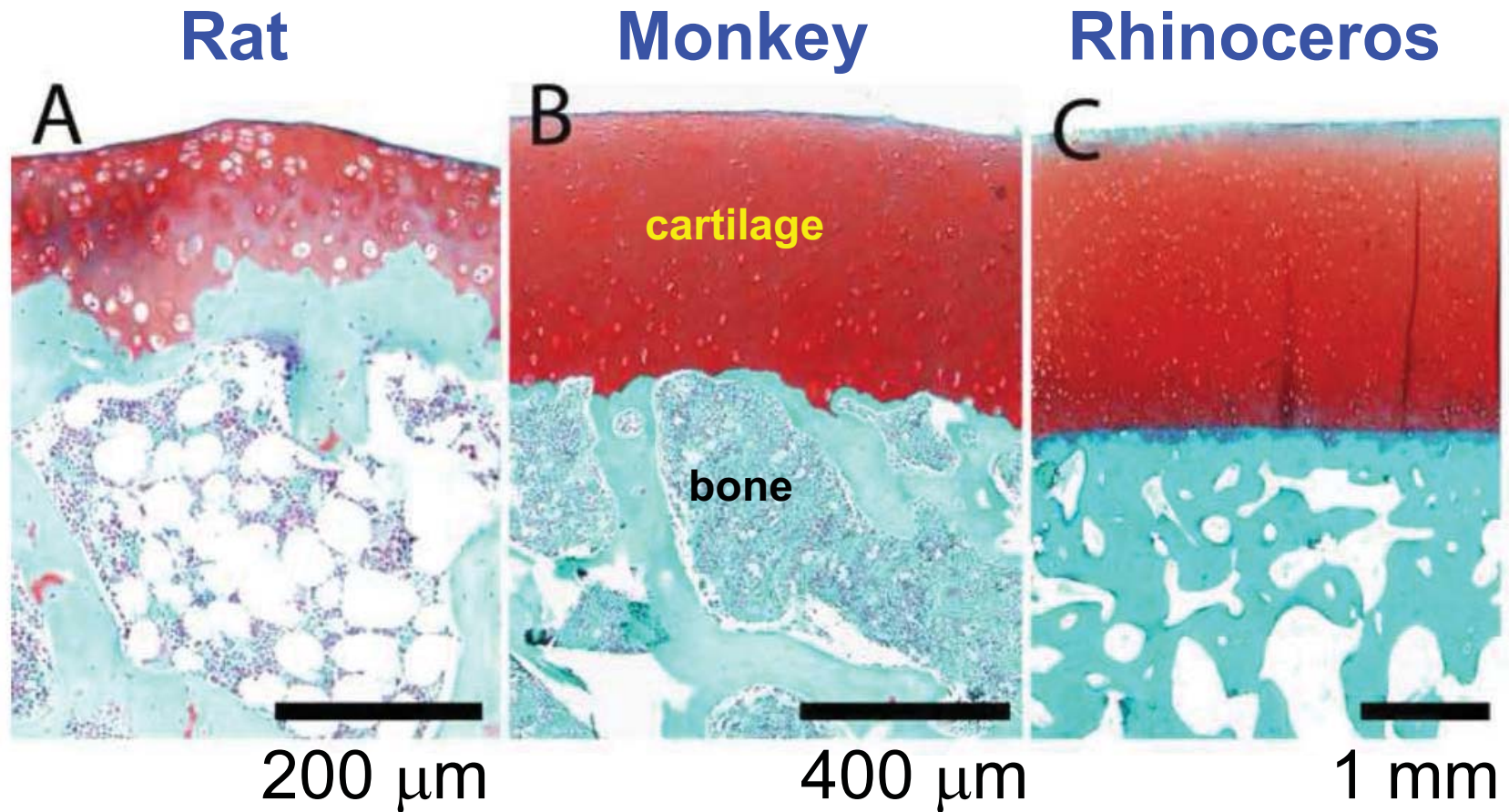
# Non-zero phase angle → Isolates Viscoelasticity of ECM (or gel, or molecular network)!!!

## Dynamic Behavior in Pure Shear



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Source: Jin, Moonsoo, and Alan J. Grodzinsky. "Effect of Electrostatic Interactions between Glycosaminoglycans on the Shear Stiffness of Cartilage: A Molecular Model and Experiments." *Macromolecules* 34, no. 23 (2001): 8330-39.

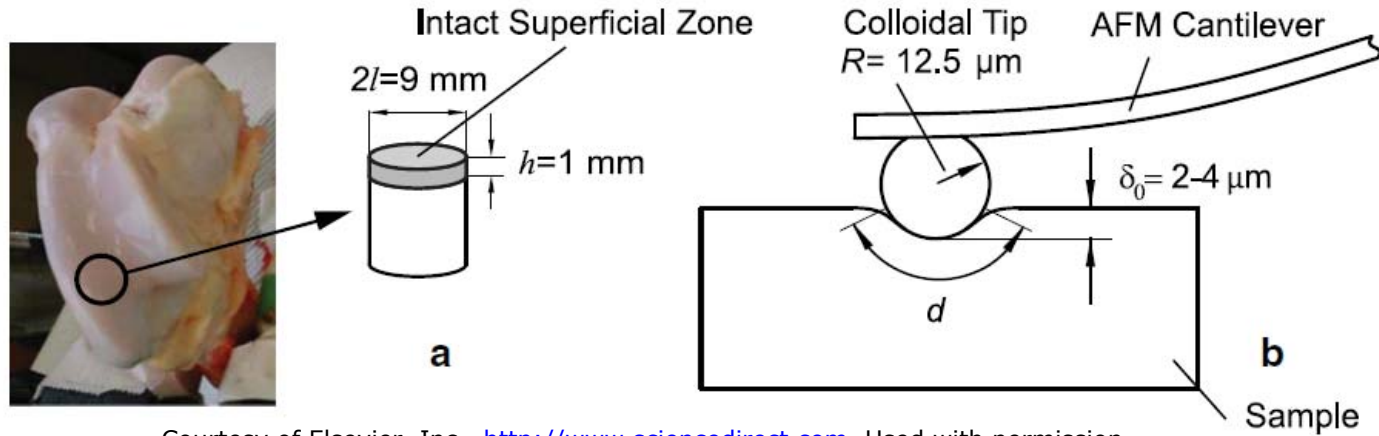
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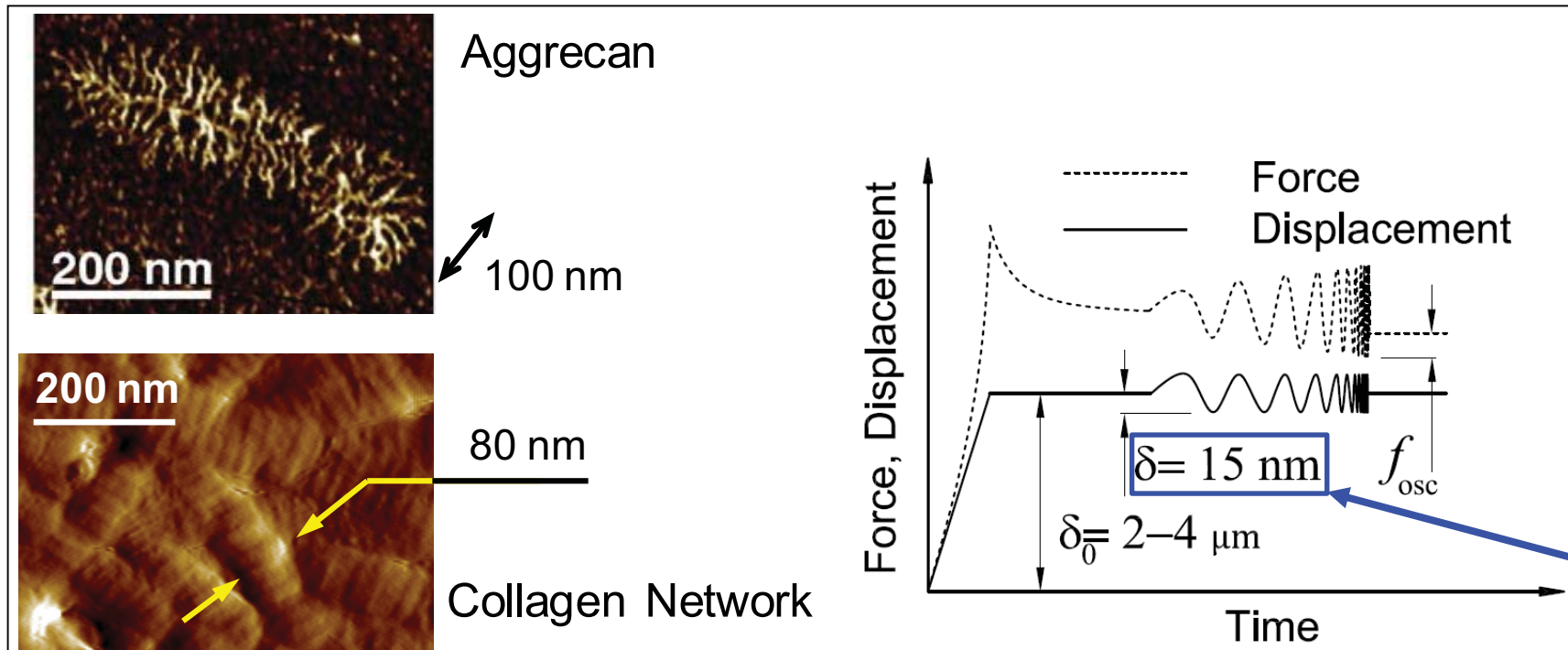
Courtesy of the authors. Used with permission.

Source: Malda, Jos, et al. "Of Mice, Men and Elephants: The Relation between Articular Cartilage Thickness and Body Mass." *306 RQH8*, no. 2 (2013): e57683.

**"Safranin-O" (red) stains Glycosaminoglycans  
(of Proteoglycans)**

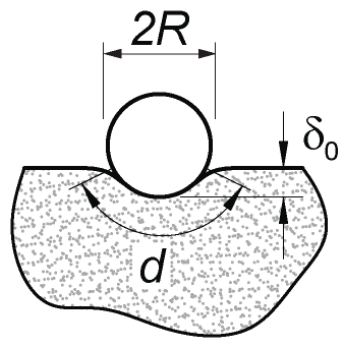
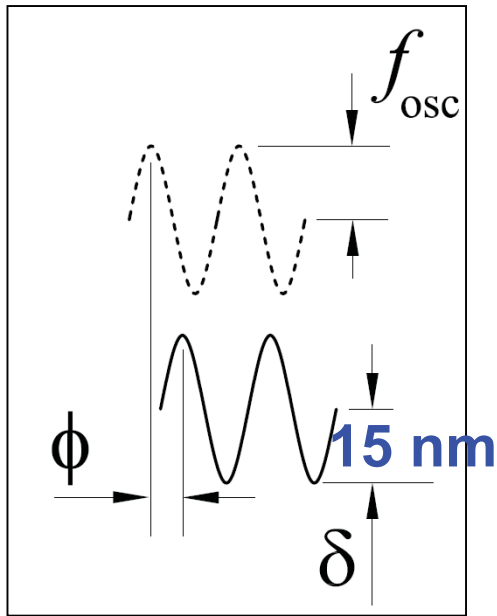


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Source: Nia, Hadi Tavakoli, et al. "Poroelectricity of Cartilage at the Nanoscale."  
*Biophysical Journal* 101, no. 9 (2011): 2304-13.

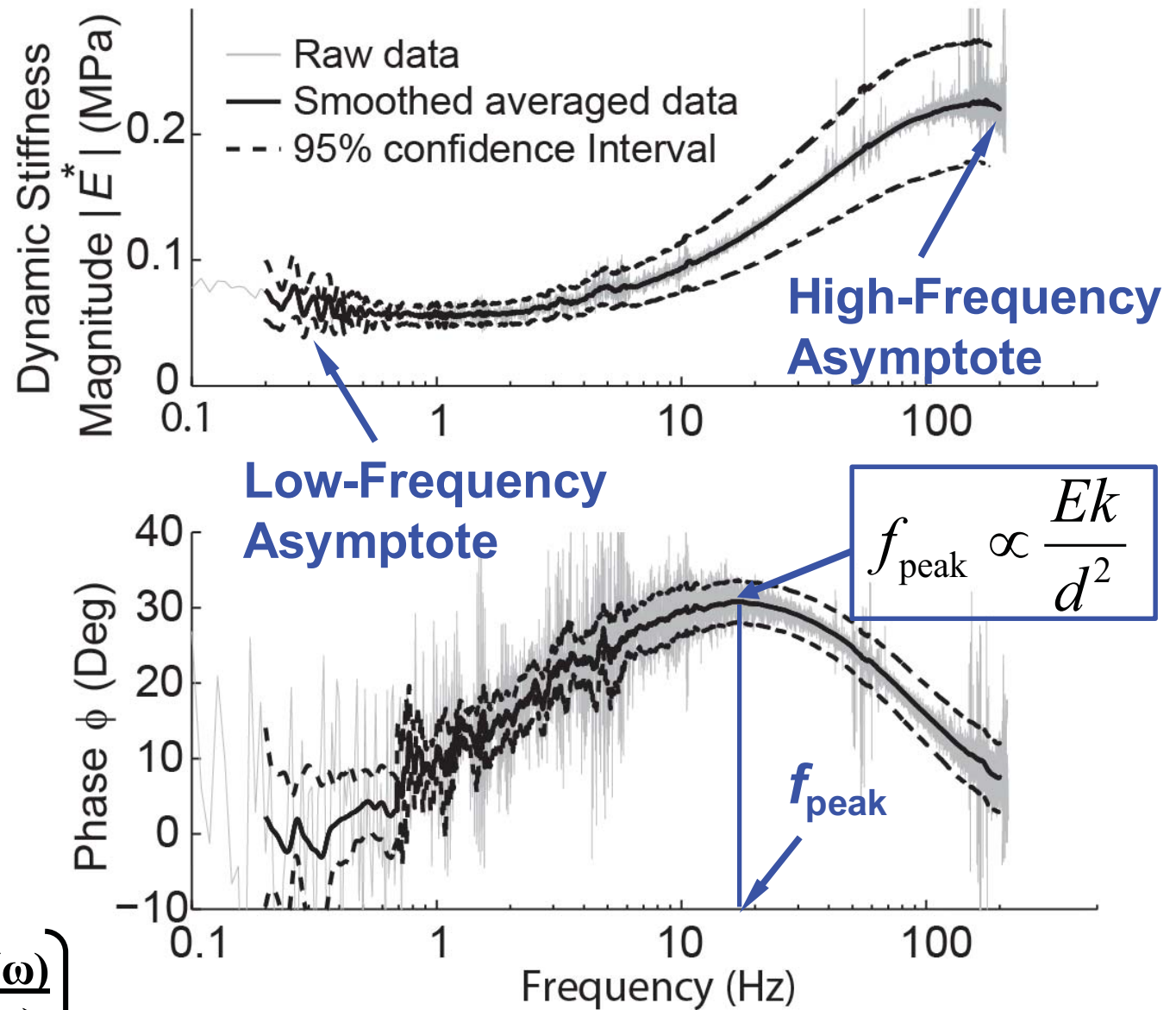


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Source: Nia, Hadi Tavakoli, et al. "High-bandwidth AFM-based Rheology Reveals that Cartilage is Most Sensitive to High Loading Rates at Early Stages of Impairment."  
*Biophysical Journal* 104, no. 7 (2013): 1529-37.

# Tissue-Level Nanomechanics: **dominated by poroelasticity**



$$\tan \phi = \left( \frac{E''(\omega)}{E'(\omega)} \right)$$



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 Source: Nia, Hadi Tavakoli, et al. "Poroelasticity of Cartilage at the Nanoscale." *Biophysical Journal* 101, no. 9 (2011): 2304-13.

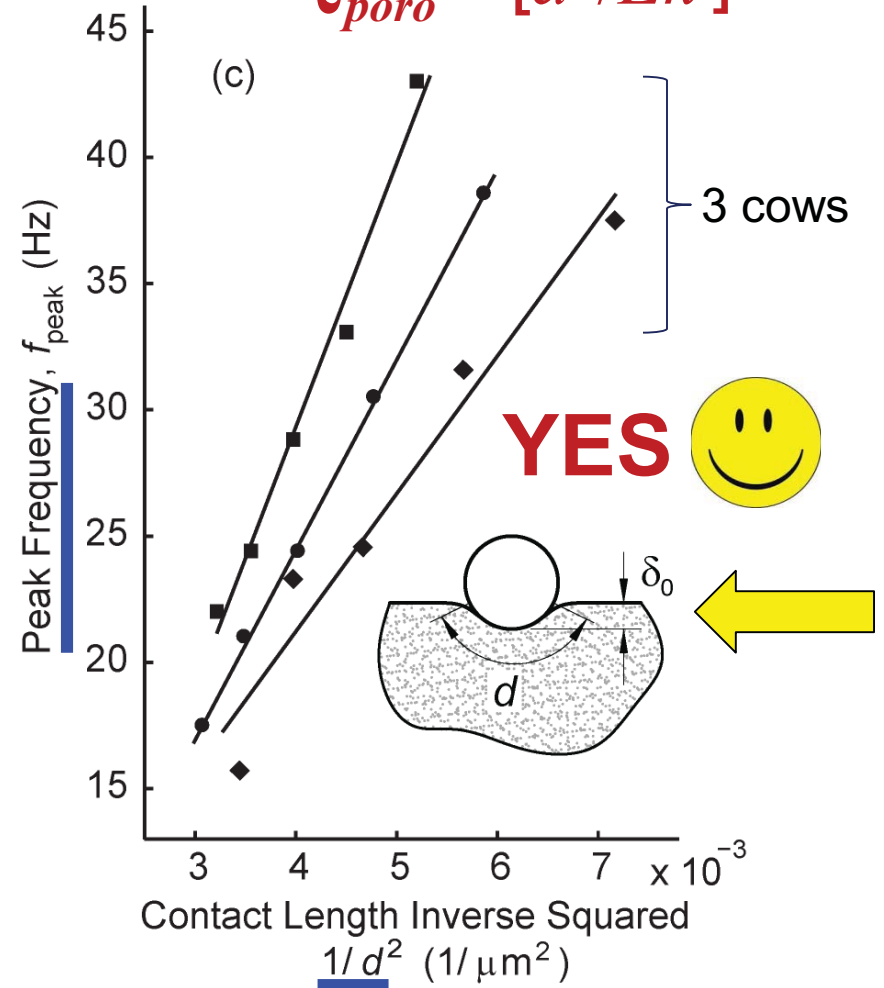
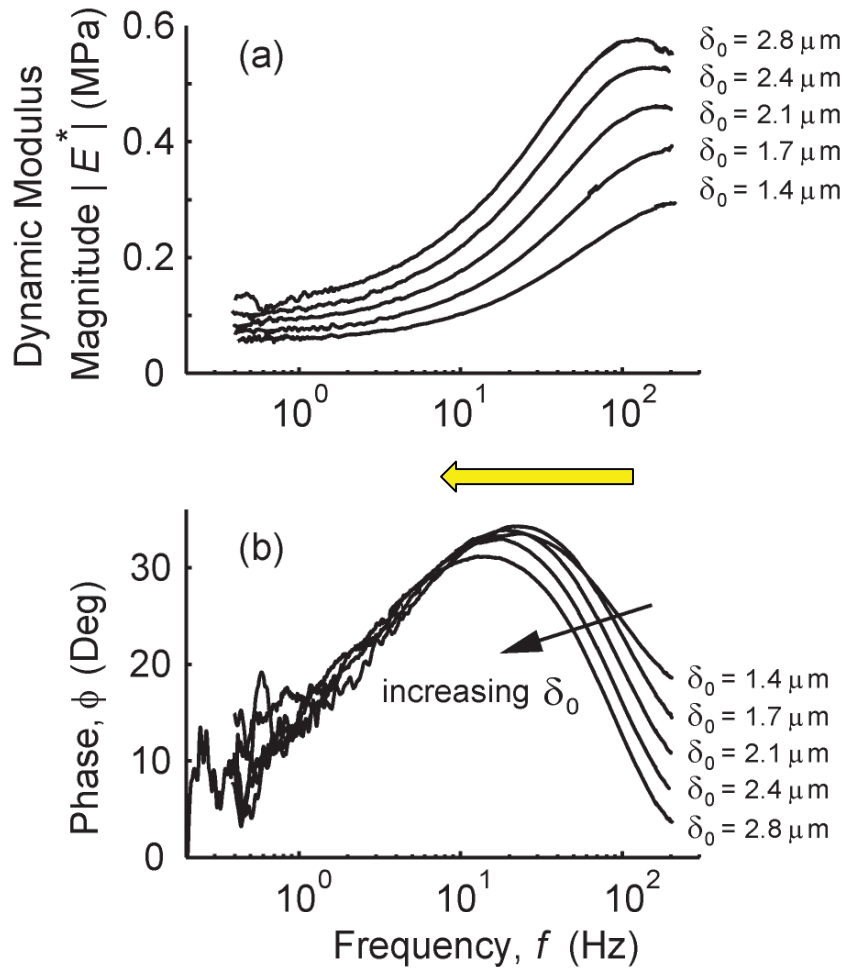
Problem removed due to copyright restrictions. See the problem in the textbook.  
Source: Grodzinsky, Alan. *Field, Forces and Flows in Biological Systems*. Garland Science, 2011.

# Is this tissue Poroelastic ??

**Test  $f_{peak}$  vs.  $d^2$**

$$f_{peak} \propto [Ek / d^2]$$

$$\tau_{poro} \propto [d^2 / Ek]$$



Nia, Biophysical J, 2011

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 Source: Nia, Hadi Tavakoli, et al. "Poroelasticity of Cartilage at the Nanoscale."  
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Spring 2015

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