

## **GENOMIC DNA PREPARATION FROM *CORYNEBACTERIUM* AND *RHODOCOCCUS*, MINI-PROTOCOL**

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1. Spin overnight culture (grown in 10ml LB) for 5 min at 6000 rpm in GSA rotor or equivalent
2. Pour off supernatant and freeze cell pellet at -20°C for 30-60 min. (or overnight at -70°C)
3. Resuspend cell pellet in 250µl fresh 10mg/ml lysozyme in TE and transfer to microcentrifuge tube
4. Add 20 µl mutanolysin (1 mg/ml)
5. Incubate at 37°C for 1-2 hr while gently shaking
6. Add 50 µl 0.5 M EDTA, 50µl 10% SDS, 50 µl 5 M NaCl; Mix gently
7. Add 10 µl fresh 20mg/ml proteinase K (or Sigma cat. no. P5568) and incubate at 37°C for 60 min.
8. Add 50 µl sodium perchlorate solution (1 g/ml) to the cell slurry; mix gently
9. Extract with one volume of phenol:chloroform:isoamyl alcohol (25:24:1); mix well; centrifuge 5 min in microcentrifuge
10. Extract aqueous phase (beware phase inversion) with 0.5 volumes of chloroform:isoamyl alcohol (24:1); mix well; centrifuge 5 min. in microcentrifuge
11. To aqueous phase add 1 volume isopropanol; microcentrifuge sample for 15 min;
12. Wash pellet; dry, resuspend DNA in 200 µl TE; may leave overnight at 4°C

For routine use, you may stop here; for higher quality DNA, proceed to step 13.

13. Add 1 µl RNase (0.5 mg/ml) and incubate at 37°C for 30 min.
14. Add 0.5 volumes 7.5M ammonium acetate
15. Extract DNA/ammonium acetate solution with one volume phenol:chloroform:isoamyl alcohol; centrifuge 5 min. in microcentrifuge
16. Extract aqueous phase with 0.5 volumes chloroform:isoamyl alcohol; centrifuge 5 min. in microcentrifuge
17. To aqueous phase add 2 volumes ethanol; microcentrifuge sample for 15 min;
18. Wash DNA in 70% ethanol, dry and resuspend in 200 µl TE.